



Physiologically-Based Pharmacokinetic Modeling of PAXLOVID™ with First-Order Absorption Kinetics

Kazuko Sagawa¹ · Jian Lin² · Rohit Jaini^{1,3} · Li Di²

Received: 16 March 2023 / Accepted: 15 May 2023 / Published online: 25 May 2023
© The Author(s), under exclusive licence to Springer Science+Business Media, LLC, part of Springer Nature 2023

Abstract

Purpose PAXLOVID™ is nirmatrelvir tablets co-packaged with ritonavir tablets. Ritonavir is used as a pharmacokinetics (PK) enhancer to reduce metabolism and increase exposure of nirmatrelvir. This is the first disclosure of Paxlovid physiologically-based pharmacokinetic (PBPK) model.

Methods Nirmatrelvir PBPK model with first-order absorption kinetics was developed using *in vitro*, preclinical, and clinical data of nirmatrelvir in the presence and absence of ritonavir. Clearance and volume of distribution were derived from nirmatrelvir PK obtained using a spray-dried dispersion (SDD) formulation where it is considered to be dosed as an oral solution, and absorption is near complete. The fraction of nirmatrelvir metabolized by CYP3A was estimated based on *in vitro* and clinical ritonavir drug-drug interaction (DDI) data. First-order absorption parameters were established for both SDD and tablet formulation using clinical data. Nirmatrelvir PBPK model was verified with both single and multiple dose human PK data, as well as DDI studies. Simcyp® first-order ritonavir compound file was also verified with additional clinical data.

Results The nirmatrelvir PBPK model described the observed PK profiles of nirmatrelvir well with predicted AUC and C_{max} values within $\pm 20\%$ of the observed. The ritonavir model performed well resulting in predicted values within twofold of observed.

Conclusions Paxlovid PBPK model developed in this study can be applied to predict PK changes in special populations, as well as model the effect of victim and perpetrator DDI. PBPK modeling continues to play a critical role in accelerating drug discovery and development of potential treatments for devastating diseases such as COVID-19. NCT05263895, NCT05129475, NCT05032950 and NCT05064800.

Keywords human pharmacokinetics · nirmatrelvir · paxlovid · PBPK modeling · ritonavir

Abbreviations

ADAM	Advanced dissolution, absorption, and metabolism	B/P	Blood-to-plasma ratio
ADME	Absorption, distribution, metabolism, and excretion	BCRP	Breast cancer resistance protein
AUC	Area under the concentration–time curve	BID	Twice a day
AUC _{inf}	Area under the concentration–time curve from time 0 to infinite	Caco-2	Human colorectal adenocarcinoma cells
		CL _{int}	Intrinsic clearance
		CL _{int,CYP3A4}	Intrinsic clearance of cytochrome P450 3A4
		CL _{po}	Oral clearance
		CL _R	Renal clearance
		C _{max}	Maximum concentration
		CYP3A	Cytochrome P450 3A
		CYP3A4	Cytochrome P450 3A4
		DDI	Drug-drug interaction
		EUA	Emergency use authorization
		f _a	Fraction absorbed
		f _m	Fraction metabolized
		f _{m,CYP3A4}	Fraction metabolized by CYP3A4
		f _u	Fraction unbound
		f _{u,gut}	Fraction unbound in enterocytes

✉ Li Di
li.di@Pfizer.com

¹ Pharmaceutical Science, Pfizer Worldwide Research and Development, 445 Eastern Point Road, Groton, CT 06340, USA

² Pharmacokinetics, Dynamics and Metabolism, Pfizer Worldwide Research and Development, 445 Eastern Point Road, Groton, CT 06340, USA

³ Pharmaceutical Science, Pfizer Worldwide Research and Development, 1 Portland Street, Cambridge, MA 02139, USA

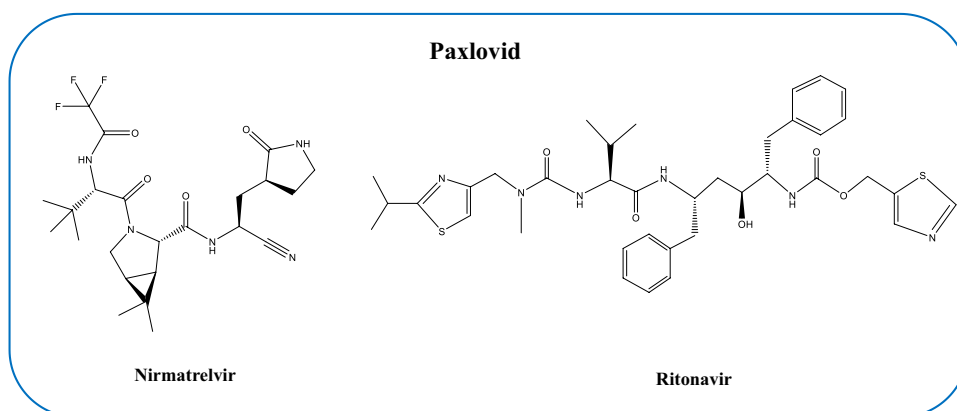
$f_{u,mic}$	Fraction unbound in microsomal incubation
f_{up}	Fraction unbound in plasma
γ	Hill coefficient
GI	Gastrointestinal
HIV	Human immunodeficiency virus
IndC ₅₀	Concentration resulting in half maximal induction
Ind _{max}	Maximal fold induction over vehicle
k_a	First-order absorption rate constant
K_{app}	The concentration of mechanism-based inhibitor associated with half maximal inactivation rate
K_i	Concentration of inhibitor that supports half maximal inhibition
k_{inact}	Maximal rate of enzyme inactivation
K_p	Tissue partition coefficient
LogP	Log10 of partition coefficient between octanol and water
MAD	Multiple ascending dose
MDCK-LE	Madin-Darby canine kidney low efflux cells
MW	Molecular weight
N	Number of subjects
NCA	Noncompartmental analysis
NMV	Nirmatrelvir
Obs	Observed
P_{app}	Apparent permeability
PBPK	Physiologically-based pharmacokinetic
P-gp	P-glycoprotein
PK	Pharmacokinetics
Pred	Predicted
Q_{gut}	A hybrid term including both villous blood flow and permeability through the enterocyte membrane
RTV	Ritonavir
SAD	Single ascending dose
SD	Single dose
SDD	Spray-dried dispersion
V_{ss}	Steady state volume of distribution

Introduction

The COVID-19 pandemic was the result of a global outbreak of coronavirus, an infectious disease caused by the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) virus. COVID-19 has resulted in more than 6.6 million death and 638 million confirmed cases globally (Coronavirus Death Toll and Trends—Worldometer (worldometers.info)). Alongside vaccines, antiviral treatment is another effective approach to combat COVID-19. PAX-LOVID™ is the first oral M^{Pro} (main protease, i.e., SARS-CoV-2 3-chymotrypsin-like cysteine protease enzyme) inhibitor that was granted emergency use authorization (EUA) in the United States, and it is currently approved or authorized for conditional or emergency use in more than 65 countries across the globe to treat high-risk COVID-19 patients. Paxlovid is nirmatrelvir tablets co-packaged with ritonavir tablets (Fig. 1). Nirmatrelvir is predominantly metabolized by CYP3A4 [1] and coadministered ritonavir inhibits CYP3A, which reduces the clearance and prolongs half-life of nirmatrelvir. This strategy has been successfully applied in various HIV treatments [2–4].

PBPK modeling and simulation has been widely used to accelerate drug discovery and development [5–7]. Analysis of historical regulatory submissions showed growing applications of PBPK and 45% of the new drug approvals containing PBPK modeling and simulation in 2019 [5, 8]. Some major applications of PBPK modeling in regulatory submission include predictions of drug-drug interactions (DDI), assessment of PK changes in special populations such as pediatrics and hepatic and renal impairment, prediction of absorption with formulation changes, food effect and others [8, 9]. This study reports a PBPK model of Paxlovid for the first time using first-order absorption kinetics that was developed and verified with *in vitro* absorption, distribution, metabolism and excretion (ADME), and clinical data, some of which have been previously published [1, 10–13]. An Advanced Dissolution, Absorption and

Fig. 1 Structure of PAX-LOVID™ – nirmatrelvir tablets co-packaged with ritonavir tablets.



Metabolism (ADAM) model for Paxlovid focusing on a more mechanistic description of nirmatrelvir dissolution and absorption will be discussed in a subsequent publication due to its complexity. The Paxlovid PBPK models developed are intended to be used to ensure product quality and desired exposure in special populations (e.g., pediatrics and pregnancy) to inform dose adjustment recommendations if necessary, as well as to predict DDI potential with comedications.

Materials and Methods

Clinical Studies

Conduct of the studies were in accordance with ethical principles derived from the Declaration of Helsinki and in compliance with International Conference on Harmonization Guidelines for Good Clinical Practice. All regulatory requirements were followed, including those affording greater protection to the safety of trial participants. The study protocols and any amendments, as well as informed consent documents were approved by the institutional review board/ethics committee. Written informed consent was obtained from all participants before any study activity. The details of the study design have been reported previously [11]. The study protocols were published in ClinicalTrials.gov with the following identifiers: NCT05263895 [relative bioavailability (Dataset #1 SDD without ritonavir, Dataset #2 SDD with ritonavir, Dataset #3 tablet with ritonavir)], NCT05129475 (food effect, Dataset #4 tablet with ritonavir), NCT05032950 (midazolam DDI, Dataset #5, tablet with ritonavir) and NCT05064800 (dabigatran etexilate DDI, Dataset #6, tablet with ritonavir). The demographic and dosing information are summarized in Table II.

PBPK Model Development and Verification

A nirmatrelvir PBPK model was developed and verified with Simcyp® version 21 release 1 (Certara, Sheffield, UK). The model was developed with *in vitro* [1], preclinical [1, 10], and human clinical data (ClinicalTrials.Org

identifiers NCT05263895, NCT05129475, NCT05032950, NCT05064800) [11, 12], where nirmatrelvir was administered with and without ritonavir. The present study focuses on model development and verification of nirmatrelvir and Paxlovid (nirmatrelvir co-dosed with ritonavir). Ritonavir compound file was developed and verified by Simcyp® using ritonavir data across multiple doses and studies [14–23]. The ritonavir first-order compound file (SV-ritonavir_FO) from the Simcyp® compound library was used for simulations without any modifications. Additional verifications of ritonavir PBPK file were performed with Paxlovid clinical data. Simcyp® midazolam (Sim-Midazolam), dabigatran etexilate (SV-Dabigatran etexilate) and dabigatran (SV-dabigatran) substrate files were used for DDI simulations without any modifications. Dabigatran etexilate substrate file was verified by Simcyp® using thirteen clinical studies. Simulations were performed with 10 trials under the fasted condition using a virtual population library of healthy volunteers in Simcyp® (Sim-Healthy Volunteers) keeping the number of subjects, age range and gender ratio consistent with the clinical trials. The scheme of nirmatrelvir model development and verification is shown in Fig. 2. A stepwise approach was taken to build and establish the first-order absorption nirmatrelvir PBPK model. Firstly, nirmatrelvir systemic parameters (clearance and volume of distribution), as well as fraction absorbed (F_a) and first-order absorption rate constant (k_a) were estimated using PK data obtained by dosing a spray dried dispersion (SDD) formulation without ritonavir. Secondly, nirmatrelvir CYP3A f_m was estimated using SDD formulation co-dosed with ritonavir. Lastly, F_a and K_a of nirmatrelvir commercial tablet was obtained using PK data of Paxlovid. The first-order absorption Paxlovid PBPK model developed was then verified using single and multiple dosing data of nirmatrelvir tablets co-administered with ritonavir, as well as both the control arms and the interactions in the DDI studies with midazolam and dabigatran etexilate.

The details of nirmatrelvir PBPK model development is discussed here. An amorphous solid dispersion formulation of nirmatrelvir, using spray drying process, was developed to enable oral solution dosing. Due to the lack of intravenous (IV) PK data, oral solution data (Dataset #1 and #2, data in Fig. 3) where healthy volunteers were dosed with

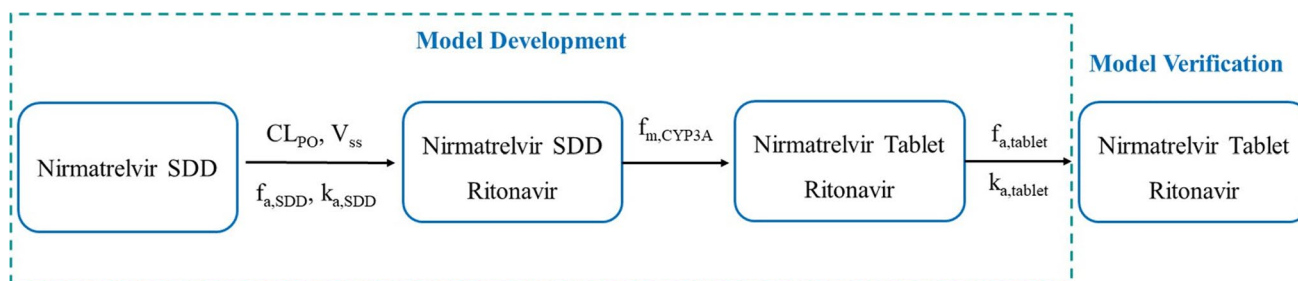
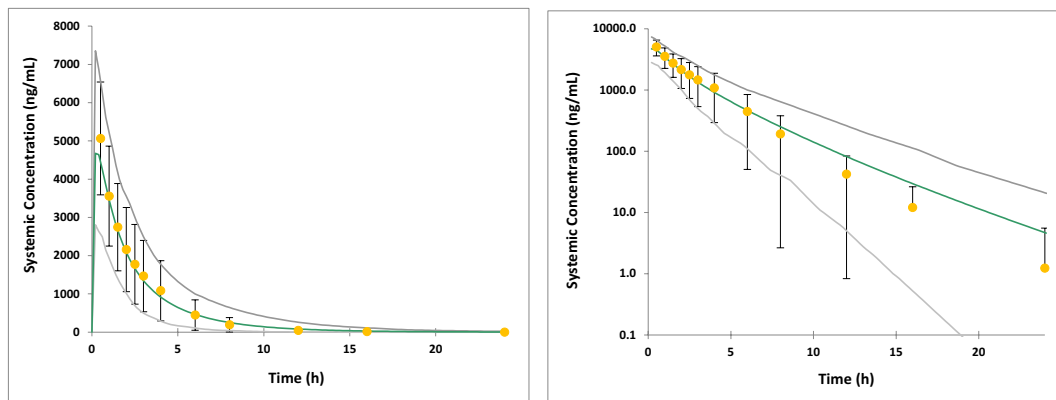


Fig. 2 Nirmatrelvir PBPK model development and verification scheme.

A Without ritonavir



B With ritonavir

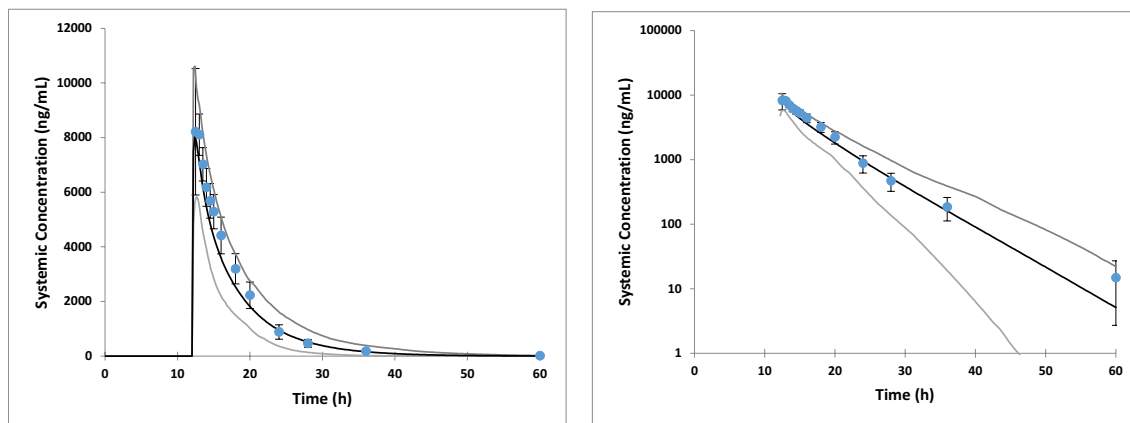


Fig. 3 Method development results: Predicted *versus* observed PK profile of nirmatrelvir SDD formulation with and without ritonavir (300 mg nirmatrelvir and three doses of 100 mg ritonavir at -12, 0, 12 h). Clinical data from NCT05263895 (relative bioavailability, Dataset #1 and #2). Human clinical data are presented as discrete points of mean value and standard deviation. Solid line in the middle is simulated mean concentration from the model. The two boundary lines represent 5th and 95th percentile. **A** Without ritonavir. **B** With ritonavir.

nirmatrelvir SDD formulation with and without co-administration of ritonavir was used for the PBPK model building to define nirmatrelvir absorption and systemic PK parameters. Clinical data with SDD formulation was treated as oral solution data. The oral clearance of nirmatrelvir ($CL_{PO} = 28.36$ L/h) and oral steady state volume of distribution ($V/F = 1$ L/kg) of nirmatrelvir were estimated from SDD solution formulation (NCT05263895, relative bioavailability, Dataset # 1, data in Fig. 3A) by fitting the clinical data. The steady state volume of distribution V_{ss} of nirmatrelvir was described using the full PBPK model in Simcyp[®] with prediction “Method 2”. “Method 2” is based upon the mechanistic approaches developed by Rodgers and Rowland [24–27], which considers partitioning of compounds into neutral lipids and phospholipids, electrostatic interactions of strong bases with acidic phospholipids, and binding to extracellular proteins of acids and bases and to lipoproteins of neutral compounds. The method calculates V_{ss} from the partition coefficient (Kp) into different tissues based on physicochemical properties (pKa , $LogD$, plasma protein binding

and blood-to-plasma ratio). The V_{ss} value was adjusted to fit the observed volume by using a global ‘ Kp scalar’ of the different tissues. Nirmatrelvir is mainly metabolized by CYP3A4 based on *in vitro* reaction phenotyping data [1]. The $f_{m,CYP3A4}$ value was optimized by using ritonavir DDI data with nirmatrelvir SDD formulation (Dataset #2, data in Fig. 3B). The intrinsic clearance by CYP3A4 ($CL_{int,CYP3A4}$) was derived using the optimized $f_{m,CYP3A4}$ value, observed oral clearance (CL_{PO}) and renal clearance with the retrograde function in Simcyp[®]. The remaining of the clearance was assigned to other metabolic clearance. Renal clearance was estimated from multiple ascending dose (MAD) study where participants received BID dosing of 75 mg, 250 mg and 750 mg of nirmatrelvir suspension with 100 mg ritonavir for 10 days [11]. Renal clearance was calculated from urinary excretion data on Day 10, and exhibited no significant dose dependence. Therefore, the observed renal clearance from the aforementioned three doses of nirmatrelvir was averaged and the mean value of 3.4 L/h was used as an input for renal clearance of nirmatrelvir in the model. Biliary

clearance of nirmatrelvir was assumed to be minimal based on studies in preclinical species [1]. Nirmatrelvir absorption was modeled using first-order absorption kinetics. The k_a for nirmatrelvir SDD tablet and commercial nirmatrelvir tablet co-packaged with ritonavir was estimated with the parameter estimation tool within Simcyp® using the observed mean plasma concentration *versus* time profiles from Dataset #1 (Data in Fig. 3A) and Dataset #3 (Data in Fig. 4), respectively. F_a of nirmatrelvir in SDD formulation was assumed to be 1 based on its solubility and absorption observed in preclinical species [1]. The f_a of nirmatrelvir from the tablet dosage form when co-administered with ritonavir was estimated using the ratio between the observed AUC_{inf} after a single dose of nirmatrelvir tablet and AUC_{inf} after a single dose of nirmatrelvir SDD formulation in presence of ritonavir [Dataset #1 (Data in Fig. 3B) and Dataset #3 (Data in Fig. 4)]. $F_{u,gut}$ was assumed to be 1 and Q_{gut} was set to 10 L/h. *In vitro* enzyme and transport inhibition/induction data were also incorporated into the model directly in order to capture any autoinhibition/autoinduction and to enable perpetrator DDI prediction. Methods of these *in vitro* DDI studies and ADME properties have been published previously [1, 28–33]. Verification of CYP3A4 autoinhibition/autoinduction of nirmatrelvir will be discussed in the ADAM model in a subsequent paper, as single and multiple ascending dose (SAD and MAD) clinical studies used suspension formulation rather than tablet and ADAM model is needed to accurately describe the disposition of nirmatrelvir in suspension dosage form. $F_{u,inc}$ values for the *in vitro* DDI parameters were estimated to be about 1 based on $f_{u,mic}$ of 0.824 at 1 mg/mL human liver microsomal protein concentration and the incubation conditions (HLM concentration of 0.01 – 0.3 mg/mL) for the *in vitro* DDI studies. Induction parameters (Ind_{max} and $IndC_{50}$) were calibrated against rifampicin.

The developed Paxlovid first-order PBPK model was verified with single and multiple dosing studies of nirmatrelvir

tablets in the presence of ritonavir tablet by comparing the AUC and C_{max} values between predicted and observed. All simulations were conducted with 10 trials maintaining the age range, proportion of females, and number of subjects consistent with the clinical studies. DDI prediction simulations of Paxlovid or ritonavir alone as inhibitors were performed with midazolam and dabigatran etexilate prodrug (dabigatran active was monitored) as substrates. Both the control arms and the interactions of DDI studies with midazolam and dabigatran etexilate were used for model verification. Model verification with a wide range of nirmatrelvir doses was conducted using the nirmatrelvir ADAM model, because suspension formulation was used for the MAD studies, and it required the ADAM model to describe the complex dissolution process of the nirmatrelvir crystalline material. The systemic parameters are identical for both the ADAM and the first-order models. The only difference between the two models is the absorption parameters. As such, the systemic parameters are considered verified with various doses by the ADAM model. Because of the complexity of the ADAM model, it will be discussed in a separate paper.

Results

The input parameters of nirmatrelvir first-order PBPK model and *in vitro* DDI values are summarized in Table I. The data sources for the input parameters of the nirmatrelvir model are also included in Table I. Information pertaining to clinical data used in model verification and model development namely, dose and dose regimen, age range of healthy volunteers, proportion of females included in the trials have been summarized in Table II. The actual clinical data are shown in Figs. 3, 4, 5 and 6 for nirmatrelvir and supplemental material Figures S1-S4 for ritonavir.

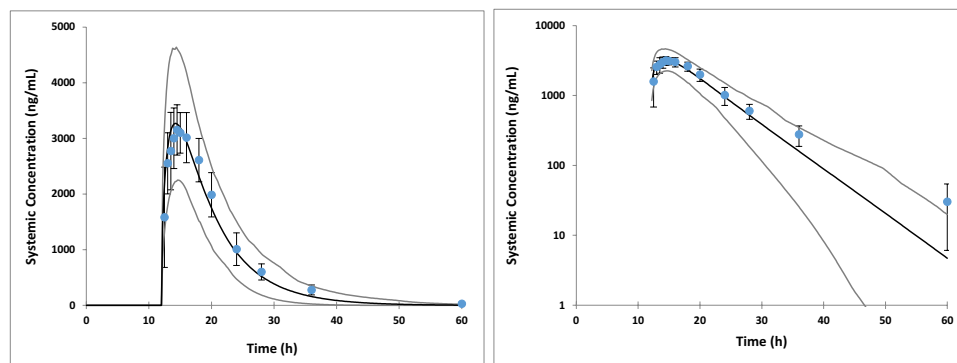


Fig. 4 Method development result: Predicted *versus* observed nirmatrelvir PK profile of Paxlovid (300 mg nirmatrelvir tablet and three doses of 100 mg ritonavir at -12, 0, 12 h). Clinical data from NCT05263895 (relative bioavailability, Dataset #3). Human clinical data are presented as discrete points of mean value and standard deviation. Solid line in the middle is simulated mean concentration from the model. The two boundary lines represent 5th and 95th percentile.

Table 1 Input Parameters of Nirmatrelvir as a Substrate in the Paxlovid PBPK Model

Category	Parameters	Value	Data Source
PhysChem Properties	MW	499.5	Calculated
	LogP	1.84	<i>In vitro</i> measurement
	Compound type	Neutral	Structure
	B/P	0.6	<i>In vitro</i> measurement
	f_{up}	0.31	<i>In vitro</i> measurement
Elimination	$CL_{int,CYP3A4}$ ($\mu\text{l}/\text{min}/\text{pmol}$)	0.148	Estimated from $f_{m,CYP3A}$ and clinical data
	Other HLM CL_{int} ($\mu\text{l}/\text{min}/\text{mg}$)	3.23	Estimated from $f_{m,CYP3A}$ and clinical data
	$f_{m,CYP3A}$	0.85	Estimated from ritonavir DDI clinical data
	$f_{u,mic}$	1	Assumed to be consistent with CL_{int}
	CL_R (L/h)	3.4	Mean clinical data from MAD
Distribution	V_{ss} (L/kg) Method 2	0.48	Estimated from SDD clinical data without ritonavir
	Kp Scalar	0.48	Estimated from SDD clinical data without ritonavir
Absorption	k_a (h^{-1}) (SDD)	2.63	Estimated from SDD clinical data without ritonavir
	f_a (SDD)	1	Estimated from SDD clinical data without ritonavir
	k_a (h^{-1}) (tablet)	0.55	Estimated from tablet clinical data with ritonavir
	f_a (tablet)	0.73	Estimated from SDD & tablet clinical data with ritonavir
	$f_{u,gut}$, Q_{gut} (L/h)	1, 10	Assumed
CYP3A4 Interaction	K_i (μM)	22.6	<i>In vitro</i> measurements
	K_{app} (μM)	13.9	
	k_{inact} (h^{-1})	0.99	
	Ind _{max} (fold)	9.74	
	IndC ₅₀ (μM)	19.04	
	γ	1.63	
Transporter Interaction	Gut Apical P-gp K_i (μM)	55.2	<i>In vitro</i> measurements
	Gut Apical OCT1 K_i (μM)	138.1	
	Liver Sinusoidal OATP1B1 K_i (μM)	44.4	
	Liver Sinusoidal OATP1B3 K_i (μM)	283.2	
	Liver Sinusoidal OCT1 K_i (μM)	138.1	
	Liver Canalicular P-gp K_i (μM)	55.2	
	Liver Canalicular MATE1 K_i (μM)	111.7	
	Kidney Apical P-gp K_i (μM)	55.2	
	Kidney Apical MATEs K_i (μM)	111.7	
	Kidney Basal OCT2 K_i (μM)	954.5	
Kidney Basal OAT3 K_i (μM)	520.6		

Model Development

Absorption

Nirmatrelvir SDD formulation of amorphous material was developed to enable solution dosing of nirmatrelvir. *In vitro* experiments confirmed that nirmatrelvir from the SDD formulation remained in solution without precipitating for a period of 24 h, significantly longer than the absorption phase *in vivo* (Pfizer data on file). The f_a of nirmatrelvir from the tablet dosage form when co-administered with ritonavir was estimated to be 0.73 using the ratio between the observed AUC_{inf} after a single dose of nirmatrelvir tablet and AUC_{inf} after a single dose of nirmatrelvir SDD formulation in

the presence of ritonavir. The estimated k_a values of nirmatrelvir tablet and SDD formulation were 0.55 h^{-1} and 2.63 h^{-1} , respectively.

Distribution

The volume of distribution of nirmatrelvir was described using full PBPK model with prediction “Method 2” within Simcyp® as described in the method section. The global Kp scalar was determined by employing the parameter estimation tool using the observed mean PK profile from the participants who received nirmatrelvir SDD formulation without ritonavir [NCT05263895 (relative bioavailability, Dataset

Table II Demographic Information of Clinical Trials and Paxlovid PBPK Model Verification Results

Dataset	N	Age (years), %female	Dosing Regimen	Nirmatrelvir Predicted/Observed Ratio	
				C_{max}	AUC
Model Development					
#1	12	35–62, 17	Nirmatrelvir SDD 300 mg single dose	NA	NA
#2	12	35–62, 17	Nirmatrelvir SDD 300 mg single dose Ritonavir 100 mg three doses at -12, 0, 12 h	NA	NA
#3	12	35–62, 17	Nirmatrelvir tablet 300 mg single dose Ritonavir 100 mg three doses at -12, 0, 12 h	NA	NA
Model Verification					
#4	12	25–73, 50	Nirmatrelvir tablet 300 mg single dose Ritonavir 100 mg three doses at -12, 0, 12 h	0.92	0.90 ^c
#5	12	21–50, 8.3	Nirmatrelvir tablet 300 mg BID 5 days ^a Ritonavir 100 mg BID 5 days ^a Midazolam 2 mg single dose on Day 5	0.98	0.95 ^d
#6	24	21–60, 62.5	Nirmatrelvir tablet 300 mg BID 2 days ^b Ritonavir 100 mg BID 2 days ^b Dabigatran etexilate 75 mg single dose on Day 2	0.93	0.95 ^d

^atotal 9 doses^btotal 3 doses^cAUC_{inf}^dAUC_τ

NA not applicable

1)]. The estimated Kp scalar was 0.48 that resulted in a predicted V_{ss} of 0.48 L/kg based on “Method 2”.

Metabolism and Renal Clearance

During the model development, initial simulations were conducted using the *in vitro* measured $f_{m,CYP3A4}$ value of 0.99. However, this did not capture the observed PK profile of nirmatrelvir when co-administered with ritonavir. Therefore, $f_{m,CYP3A4}$ was optimized using the observed PK profile from participants who received 300 mg nirmatrelvir SDD

formulation with three doses of 100 mg ritonavir at -12 h, 0 h and 12 h [NCT05263895 (relative bioavailability, Dataset #2)]. The sensitivity analysis was performed using an increment of ~0.05 with three $f_{m,CYP3A4}$ values at 0.80, 0.85 and 0.99 (Supplemental Material Table S1). An $f_{m,CYP3A4}$ of 0.85 best described the PK profile of nirmatrelvir in the presence of ritonavir with the predicted/observed ratios of 0.92 and 0.87 for C_{max} and AUC_{inf}, respectively (Supplemental Material Table S1). The current model with an $f_{m,CYP3A4}$ of 0.85 described the available clinical data well without any biases. More precise estimate of f_m can lead to overfitting of

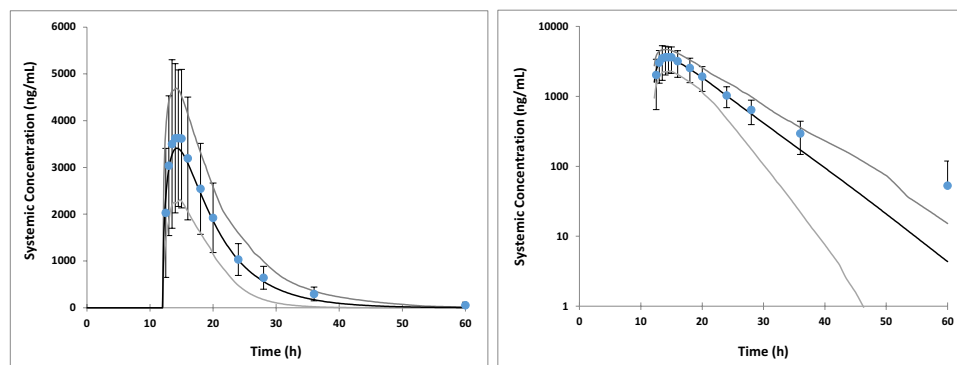


Fig. 5 Method Verification Result: Predicted *versus* observed nirmatrelvir PK profile of Paxlovid (300 mg nirmatrelvir tablets and three doses of 100 mg ritonavir at -12, 0, 12 h). Clinical data from NCT05129475 (food effect, Dataset #4). Human clinical data are presented as discrete points of mean value and standard deviation. Solid line in the middle is simulated mean concentration from the model. The two boundary lines represent 5th and 95th percentile.

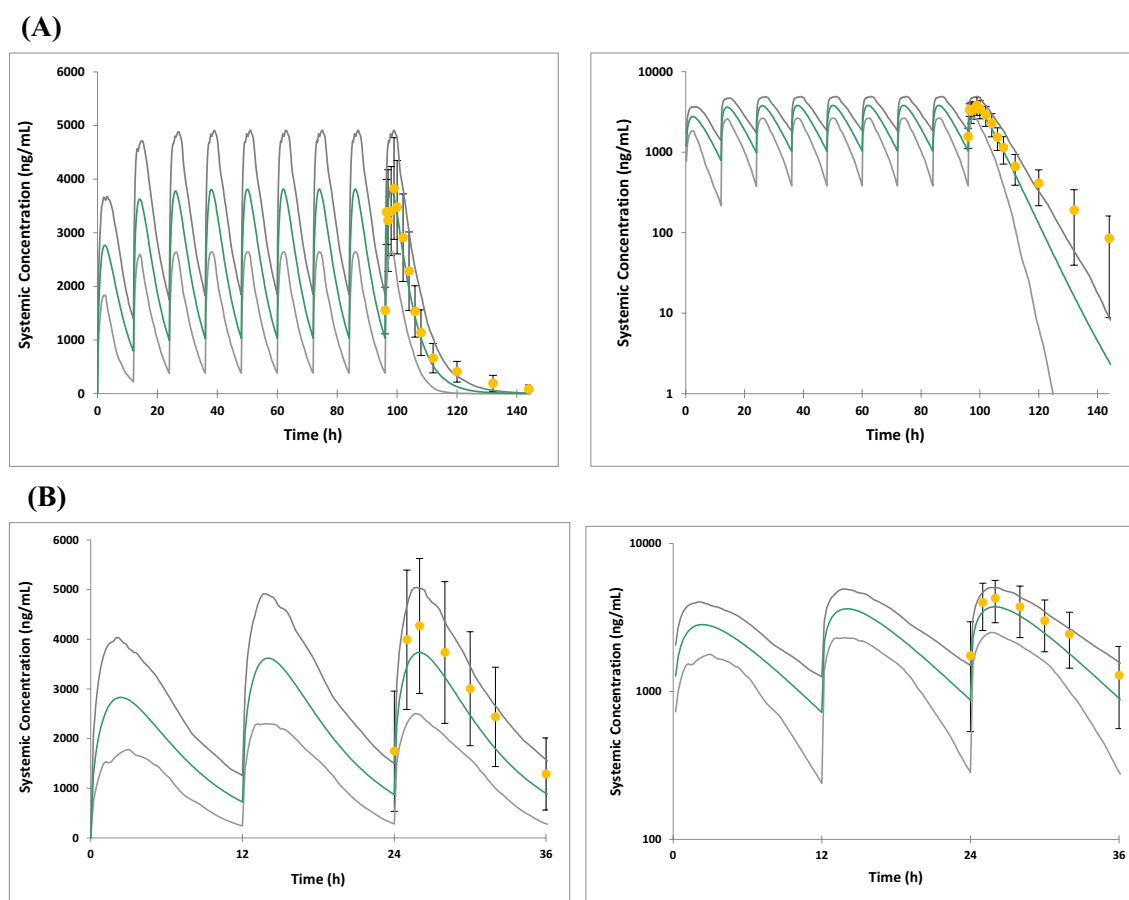


Fig. 6 Method Verification Result: Predicted *versus* observed nirmatrelvir PK profile of Paxlovid: **(A)** 300 mg nirmatrelvir tablets and 100 mg ritonavir BID for 5 days, 9 doses total. Clinical data from NCT05032950 (midazolam DDI, Dataset #5). **(B)** 300 mg nirmatrelvir tablets and 100 mg ritonavir BID for 2 days, 3 doses total. Clinical data from NCT05064800 (dabigatran etexilate DDI, Dataset #6). Human clinical data are presented as discrete points of mean value and standard deviation. Solid line in the middle is simulated mean concentration from the model. The two boundary lines represent 5th and 95th percentile.

the data considering interindividual variability. $CL_{int,CYP3A4}$ was estimated to be $0.148 \mu\text{l}/\text{min}/\text{pmol}$ using the retrograde function in Simcyp[®] with the optimized $f_{m,CYP3A4}$ of 0.85, observed geometric mean of CL_{po} (28.36 L/h), renal clearance of 3.4 L/h (clinical data), and F_a of 1. The remaining f_m of 0.15 was assigned to additional HLM CL_{int} (3.23 $\mu\text{l}/\text{min}/\text{mg}$). Renal clearance of 3.4 L/h was obtained from clinical data and entered into nirmatrelvir compound file (see Materials and Methods section). Nirmatrelvir renal clearance did not change in the presence and absence of ritonavir.

Model Development Results

The systemic parameters (clearance and volume of distribution) for the nirmatrelvir PBPK model was successfully developed using data from studies in NCT05263895 (relative bioavailability, Dataset #1 and #2) with SDD formulation (modeled as oral solution). These systemic parameters determined with SDD formulation were used to estimate k_a for the nirmatrelvir tablet formulation. The model described

the observed clinical PK of nirmatrelvir well for both of the formulations with and without ritonavir (Figs. 3 and 4). In addition, the Simcyp[®] ritonavir compound file with first-order absorption kinetics adequately described the observed ritonavir plasma concentration profiles (Supplemental Material, Table S2 and Figures S1-S2).

Model Verification

The established Paxlovid PBPK model was verified with data from three single and multiple dose clinical studies for food effect and DDI [NCT05129475 (food effect, Dataset #4), NCT05032950 (midazolam DDI, Dataset #5) and NCT05064800 (dabigatran etexilate DDI, Dataset #6)]. The verification results are shown in Table II and Figs. 5 and 6. The model successfully predicted the nirmatrelvir exposures with predicted/observed ratios ranging from 0.90 to 0.98 for AUC_{inf} and C_{max} (Table II). This indicates that the estimated model input parameters were appropriate to predict the nirmatrelvir exposures. The fed state arm of the clinical

data was not used in the model verification as it requires the use of the ADAM model, which will be discussed in a subsequent publication. DDI clinical studies with midazolam (CYP3A substrate) and dabigatran etexilate (P-gp substrate) were conducted with both Paxlovid and ritonavir alone as perpetrators for comparison purpose (Table III). Given that the C_{max} and AUC_{last} ratios as a result of DDI were similar between Paxlovid and ritonavir alone, it was concluded that perpetrator DDIs of Paxlovid for CYP3A and P-gp were mainly driven by ritonavir rather than nirmatrelvir. The verifications of Simcyp[®] ritonavir compound file using these studies are summarized in the Supplemental Material (Table S2 and Figures S3–S4). Simcyp[®] ritonavir compound file described the clinical data well for both PK and DDI with predictions falling within twofold of the observed values (Table III and Supplemental Material Table S2).

Discussion

Nirmatrelvir first-order absorption PBPK model was successfully developed by characterizing the ADME properties of nirmatrelvir and its interaction with ritonavir. The model was developed with nirmatrelvir SDD formulation and tablet with and without ritonavir, and verified with all relevant clinical data including single and multiple dose studies. Certain clinical data were not included in the model verification, such as the SAD and MAD studies with nirmatrelvir suspension, due to complex absorption processes that require ADAM model to mechanistically describe the dissolution/absorption properties, which will be discussed in a subsequent publication.

As a peptide mimetic, nirmatrelvir has many unique ADME properties leading to its PK behavior. Nirmatrelvir has low passive permeability in the MDCK-LE [34] (P_{app} 1.76×10^{-6} cm/s) and Caco-2 (P_{app} $0.66\text{--}1.19 \times 10^{-6}$ cm/s) *in vitro* assays [1]. Despite the low passive permeability *in*

vitro, nirmatrelvir is well absorbed in preclinical species (e.g., rat) [1] and humans. The *in vitro-in vivo* disconnect on nirmatrelvir oral absorption is currently not well-understood, although similar observations have been reported in the literature [35]. Nirmatrelvir is a P-gp substrate, but not a BCRP substrate [1]. P-gp efflux doesn't appear to have significant impact on oral absorption of nirmatrelvir at the doses studied as SDD formulation showed near complete absorption, and no significant supra-proportional exposure was observed with increased dose in rat [36]. This is consistent with literature reports of minimal impact of P-gp efflux on oral absorption for compounds with relatively high solubility and permeability [37–43].

In vitro $f_{m,CYP3A4}$ of nirmatrelvir was estimated to be 0.99 based on formation rate of the major metabolites with selective inhibitors [1]. *In vivo* $f_{m,CYP3A4}$ was estimated to be 0.85 based on ritonavir DDI study, indicating some uncertainties of *in vitro* assays in estimating *in vivo* f_m . Sensitivity analysis of f_m can help further understand the victim DDI risk *in vivo*. With ritonavir coadministration, the major clearance pathway of nirmatrelvir shifted from CYP3A metabolism to renal elimination [12], with an increased half-life from 2 to 7 hour. At 100 mg dose, ritonavir completely inhibits CYP3A [18]. Ritonavir at 100 mg dose is commonly used as a booster for protease inhibitors in HIV therapy (e.g., in combination with lopinavir, saquinavir, or atazanavir) [4]. Compared to ritonavir, nirmatrelvir is a much weaker CYP3A and P-gp inhibitor based on *in vitro* inhibition potency. As such, Paxlovid perpetrator DDIs for CYP3A and P-gp are mainly driven by ritonavir. The slight over-prediction of midazolam DDI might be due to variability of the clinical data. The mean and 90% confidence interval of $AUC_{last}R$ are 14.5 (12.2, 17.2) for Paxlovid and 16.8 (14.1, 19.9) for ritonavir DDI with midazolam. An $AUC_{\infty}R$ of 23.8 was reported for ritonavir (100 mg BID) with midazolam DDI in the literature [17]. The underprediction of dabigatran etexilate DDI is likely because the *in vitro* K_i for P-gp might be

Table III Paxlovid and Ritonavir Drug-Drug Interactions with Midazolam and Dabigatran Etexilate

Dataset	Perpetrator	Object	Observed		Predicted		Predicted/Observed	
			$C_{max}R$	$AUC_{last}R$	$C_{max}R$	$AUC_{last}R$	$C_{max}R$	$AUC_{last}R$
#5	Paxlovid ^a	Midazolam ^a	3.68	14.5	4.36	24.7	1.18	1.70
	Ritonavir ^b	Midazolam ^b	3.87	16.8	4.40	25.3	1.14	1.51
#6	Paxlovid ^c	Dabigatran Etexilate ^c	2.33	2.15	1.28	1.28	0.549	0.595
	Ritonavir ^d	Dabigatran Etexilate ^d	1.72	1.60	1.24	1.24	0.721	0.775

^a300 mg nirmatrelvir and 100 mg ritonavir BID for 5 days (9 doses total), midazolam 2 mg single dose on Day 5

^b100 mg ritonavir BID for 5 days (9 doses total), midazolam 2 mg single dose on Day 5

^c300 mg nirmatrelvir and 100 mg ritonavir BID for 2 days (3 doses total), dabigatran etexilate 75 mg single dose on Day 2

^d100 mg ritonavir BID for 2 days (3 doses total), dabigatran etexilate 75 mg single dose on Day 2

higher than the *in vivo* K_i . Scaling factors are usually needed to more accurately predict *in vivo* transporter-mediated DDI. For perpetrator DDI predictions, sensitivity analysis around K_i values is recommended to account for the uncertainties in experimental K_i values for *in vivo* translation [44–49]. When co-administered with ritonavir, CYP3A4 metabolism of nirmatrelvir is inhibited leading to increased fractional renal clearance ($f_{CL,R}$). Although nirmatrelvir renal clearance (CL_R) remains the same in the presence and absence of ritonavir, the percentage of nirmatrelvir that is eliminated renally significantly increases when co-administered with ritonavir. Consequently, a reduced nirmatrelvir dose is recommended for moderate renal impairment patients, and Paxlovid is not recommended in patients with severe renal impairment (Paxlovid HCP FS 09,262,022 (fda.gov)).

The first-order absorption PBPK model of Paxlovid has been applied to predict the DDI risk with weak and moderate CYP3A inducers and suggested no significant risk of DDI (Supplemental Material, Table S3). These PBPK simulation results were accepted by FDA. In addition, the Paxlovid models are being applied to inform dose selection of special populations (e.g., organ impairments, pediatrics, and pregnancy), DDI predictions and bioequivalence simulations.

Conclusions

A Paxlovid PBPK model with first-order absorption kinetics has been successfully developed and verified. The model describes the nirmatrelvir PK well with and without ritonavir from multiple clinical studies using tablet formulation. The PBPK model not only helps to understand Paxlovid disposition mechanically, but also enables forward predictions of DDI with co-meds, exposure changes in special populations to inform dose adjustment recommendation, and support product label. The Paxlovid PBPK model enables informed internal decisions and regulatory submissions to address various queries. PBPK modeling continues to play a pivotal role in accelerating drug discovery and development of potentially life-saving medicines as has been described herein for Paxlovid.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s11095-023-03538-5>.

Acknowledgements Authors greatly appreciate the help of Demetria Baker DeBartolo in data QC and report formatting; thanks to Rajendra S Kadam and Donna S. Cox for managing the clinical studies; thanks to many Pfizer colleagues for their input and discussion.

Funding This study was sponsored by Pfizer Inc., New York, NY, USA.

Data Availability Upon request, and subject to review, Pfizer will provide the data that support the findings of this study. Subject to certain criteria, conditions and exceptions, Pfizer may also provide access

to the related individual de-identified participant data. See <https://www.pfizer.com/science/clinical-trials/trial-data-and-results> for more information.

Declarations

Conflict of Interest All authors are employees of Pfizer Inc., New York, NY, USA and may hold stock or stock options.

References

- Eng H, Dantonio AL, Kadar EP, Obach RS, Di L, Lin J, Patel NC, Boras B, Walker GS, Novak JJ, Kimoto E, Singh RSP, Kalgutkar AS. Disposition of nirmatrelvir, an orally bioavailable inhibitor of SARS-CoV-2 3C-like protease, across animals and humans. *Drug Metab Dispos.* 2022;50(5):576–90.
- Cooper CL, van Heeswijk RPG, Gallicano K, Cameron DW. A review of low-dose ritonavir in protease inhibitor combination therapy. *Clin Infect Dis.* 2003;36(12):1585–92.
- De Nicolo A, Simiele M, Calcagno A, Abdi AM, Bonora S, Di Perri G, D'Avolio A. Intracellular antiviral activity of low-dose ritonavir in boosted protease inhibitor regimens. *Antimicrob Agents Chemother.* 2014;58(7):4042–7.
- Hull MW, Montaner JSG. Ritonavir-boosted protease inhibitors in HIV therapy. *Ann Med.* 2011;43(5):375–88.
- Chu X, Prasad B, Neuhoff S, Yoshida K, Leeder JS, Mukherjee D, Taskar K, Varma MVS, Zhang X, Yang X, Galetin A. Clinical Implications of Altered Drug Transporter Abundance/Function and PBPK Modeling in Specific Populations: An ITC Perspective. *Clin Pharmacol Ther.* 2022;112(3):501–26.
- Shebley M, Sandhu P, Emami Riedmaier A, Jamei M, Narayanan R, Patel A, Peters SA, Reddy VP, Zheng M, de Zwart L, Beneton M, Bouzom F, Chen J, Chen Y, Cleary Y, Collins C, Dickinson GL, Djebli N, Einolf HJ, Gardner I, Huth F, Kazmi F, Khalil F, Lin J, Odinecs A, Patel C, Rong H, Schuck E, Sharma P, Wu SP, Xu Y, Yamazaki S, Yoshida K, Rowland M. Physiologically Based Pharmacokinetic Model Qualification and Reporting Procedures for Regulatory Submissions: A Consortium Perspective. *Clin Pharmacol Ther.* 2018;104(1):88–110.
- Grimstein M, Yang Y, Zhang X, Grillo J, Huang SM, Zineh I, Wang Y. Physiologically Based Pharmacokinetic Modeling in Regulatory Science: An Update From the U.S. Food and Drug Administration's Office of Clinical Pharmacology. *J Pharm Sci.* 2019;108(1):21–5.
- Zhang X, Yang Y, Grimstein M, Fan J, Grillo JA, Huang S-M, Zhu H, Wang Y. Application of PBPK Modeling and Simulation for Regulatory Decision Making and Its Impact on US Prescribing Information: An Update on the 2018–2019 Submissions to the US FDA's Office of Clinical Pharmacology. *J Clin Pharmacol.* 2020;60(S1):S160–78.
- Grimstein M, Yang Y, Zhang X, Grillo J, Huang S-M, Zineh I, Wang Y. Physiologically Based Pharmacokinetic Modeling in Regulatory Science: An Update From the U.S. Food and Drug Administration's Office of Clinical Pharmacology. *J Pharm Sci.* 2019;108(1):21–5.
- Owen DR, Allerton CMN, Anderson AS, Aschenbrenner L, Avery M, Berritt S, Boras B, Cardin RD, Carlo A, Coffman KJ, Dantonio A, Di L, Eng H, Ferre R, Gajiwala KS, Gibson SA, Greasley SE, Hurst BL, Kadar EP, Kalgutkar AS, Lee JC, Lee J, Liu W, Mason SW, Noell S, Novak JJ, Obach RS, Ogilvie K, Patel NC, Pettersson M, Rai DK, Reese MR, Sammons MF, Sathish JG, Singh RSP, Stepan CM, Stewart AE, Tuttle JB, Updyke L, Verhoest PR, Wei L, Yang Q, Zhu Y. An oral

- SARS-CoV-2 Mpro inhibitor clinical candidate for the treatment of COVID-19. *Science*. 2021;374(6575):1586–93.
11. Singh RSP, Toussi SS, Hackman F, Chan PL, Rao R, Allen R, Van Eyck L, Pawlak S, Kadar EP, Clark F, Shi H, Anderson AS, Binks M, Menon S, Nucci G, Bergman A. Innovative Randomized Phase I Study and Dosing Regimen Selection to Accelerate and Inform Pivotal COVID-19 Trial of Nirmatrelvir. *Clin Pharmacol Ther*. 2022;112(1):101–11.
 12. Singh RSP, Walker GS, Kadar EP, Cox LM, Eng H, Sharma R, et al. Metabolism and excretion of nirmatrelvir in humans using quantitative fluorine nuclear magnetic resonance spectroscopy: a novel approach for accelerating drug development. *Clin Pharmacol Ther*. 2022;112(6):1201–6.
 13. Hammond J, Leister-Tebbe H, Gardner A, Abreu P, Bao W, Wisemandle W, Baniecki M, Hendrick VM, Damle B, Simon-Campos A, Pypstra R, Rusnak JM. Oral nirmatrelvir for high-risk, nonhospitalized adults with Covid-19. *N Engl J Med*. 2022;386(15):1397–408.
 14. Eichbaum C, Cortese M, Blank A, Burhenne J, Mikus G. Concentration effect relationship of CYP3A inhibition by ritonavir in humans. *Eur J Clin Pharmacol*. 2013;69(10):1795–800.
 15. Greenblatt DJ, Peters DE, Oleson LE, Harmatz JS, MacNab MW, Berkowitz N, Zinny MA, Court MH. Inhibition of oral midazolam clearance by boosting doses of ritonavir, and by 4,4-dimethyl-benziso-(2H)-selenazine (ALT-2074), an experimental catalytic mimic of glutathione oxidase. *Br J Clin Pharmacol*. 2009;68(6):920–7.
 16. Ieiri I, Tsunemitsu S, Maeda K, Ando Y, Izumi N, Kimura M, Yamane N, Okuzono T, Morishita M, Kotani N, Kanda E, Deguchi M, Matsuguma K, Matsuki S, Hirota T, Irie S, Kusuhara H, Sugiyama Y. Mechanisms of pharmacokinetic enhancement between ritonavir and saquinavir; micro/small dosing tests using midazolam (CYP3A4), fexofenadine (p-glycoprotein), and pravastatin (OATP1B1) as probe drugs. *J Clin Pharmacol*. 2013;53(6):654–61.
 17. Mathias AA, German P, Murray BP, Wei L, Jain A, West S, Warren D, Hui J, Kearney BP. Pharmacokinetics and pharmacodynamics of GS-9350: a novel pharmacokinetic enhancer without anti-HIV activity. *Clin Pharmacol Ther*. 2010;87(3):322–9.
 18. Mathias AA, West S, Hui J, Kearney BP. Dose-response of ritonavir on hepatic CYP3A activity and elvitegravir oral exposure. *Clin Pharmacol Ther*. 2009;85(1):64–70.
 19. Ouellet D, Hsu A, Granneman GR, Carlson G, Cavanaugh J, Guenther H, Leonard JM. Pharmacokinetic interaction between ritonavir and clarithromycin. *Clin Pharmacol Ther*. 1998;64(4):355–62.
 20. Greenblatt DJ, von Moltke LL, Harmatz JS, Durol AL, Daily JP, Graf JA, Mertzanis P, Hoffman JL, Shader RI. Alprazolam-ritonavir interaction: implications for product labeling. *Clin Pharmacol Ther*. 2000;67(4):335–41.
 21. Kirby BJ, Collier AC, Kharasch ED, Dixit V, Desai P, Whittington D, Thummel KE, Unadkat JD. Complex drug interactions of HIV protease inhibitors 2: in vivo induction and in vitro to in vivo correlation of induction of cytochrome P450 1A2, 2B6, and 2C9 by ritonavir or nelfinavir. *Drug Metab Dispos*. 2011;39(12):2329–37.
 22. Kirby BJ, Collier AC, Kharasch ED, Whittington D, Thummel KE, Unadkat JD. Complex drug interactions of the HIV protease inhibitors 3: effect of simultaneous or staggered dosing of digoxin and ritonavir, nelfinavir, rifampin, or bupropion. *Drug Metab Dispos*. 2012;40(3):610–6.
 23. Aarnoutse RE, Kleinnijenhuis J, Koopmans PP, Touw DJ, Wieling J, Hekster YA, Burger DM. Effect of low-dose ritonavir (100 mg twice daily) on the activity of cytochrome P450 2D6 in healthy volunteers. *Clin Pharmacol Ther*. 2005;78(6):664–74.
 24. Rodgers T, Rowland M. Mechanistic approaches to volume of distribution predictions: understanding the processes. *Pharm Res*. 2007;24(5):918–33.
 25. Rodgers T, Rowland M. Physiologically based pharmacokinetic modelling 2: predicting the tissue distribution of acids, very weak bases, neutrals and zwitterions. *J Pharm Sci*. 2006;95(6):1238–57.
 26. Rodgers T, Leahy D, Rowland M. Physiologically Based Pharmacokinetic Modeling 1: Predicting the Tissue Distribution of Moderate-to-Strong Bases. *J Pharm Sci*. 2005;94(6):1259–76.
 27. Rodgers T, Rowland M, Rodgers T, Rowland M. Physiologically-based pharmacokinetic modeling 2: predicting the tissue distribution of acids, very weak bases, neutrals and zwitterions. *J Pharm Sci*. 2006;95:1238–57.
 28. Lazzaro S, West MA, Eatemadpour S, Feng B, Varma MVS, Rodrigues AD, et al. Translatability of in vitro inhibition potency to in vivo P-glycoprotein mediated drug interaction risk. *J Pharm Sci*. 2023;112(6):1715–23.
 29. Costales C, Lin J, Kimoto E, Yamazaki S, Gosset JR, Rodrigues AD, Lazzaro S, West MA, West M, Varma MVS. Quantitative prediction of breast cancer resistant protein mediated drug-drug interactions using physiologically-based pharmacokinetic modeling. *CPT: Pharmacometrics Syst Pharmacol*. 2021;10(9):1018–31.
 30. Kimoto E, Costales C, West MA, Bi YA, Vourvahis M, David Rodrigues A, Varma MVS. Biomarker-Informed Model-Based Risk Assessment of Organic Anion Transporting Polypeptide 1B Mediated Drug-Drug Interactions. *Clin Pharmacol Ther*. 2022;111(2):404–15.
 31. Mathialagan S, Feng B, Rodrigues AD, Varma MVS. Drug-Drug Interactions Involving Renal OCT2/MATE Transporters: Clinical Risk Assessment May Require Endogenous Biomarker-Informed Approach. *Clin Pharmacol Ther*. 2021;110(4):855–9.
 32. Feng B, Hurst S, Lu Y, Varma MV, Rotter CJ, El-Kattan A, Lockwood P, Corrigan B. Quantitative prediction of renal transporter-mediated clinical drug-drug interactions. *Mol Pharm*. 2013;10(11):4207–15.
 33. Ryu S, Woody N, Chang G, Mathialagan S, Varma MVS. Identification of Organic Anion Transporter 2 Inhibitors: Screening, Structure-Based Analysis, and Clinical Drug Interaction Risk Assessment. *J Med Chem*. 2022;65(21):14578–88.
 34. Di L, Whitney-Pickett C, Umland JP, Zhang H, Zhang X, Gebhard DF, Lai Y, Federico JJ, Davidson RE, Smith R, Reyner EL, Lee C, Feng B, Rotter C, Varma MV, Kempshall S, Fenner K, El-kattan AF, Liston TE, Troutman MD. Development of a new permeability assay using low-efflux MDCKII cells. *J Pharm Sci*. 2011;100(11):4974–85.
 35. Price E, Kalvass JC, DeGoey D, Hosmane B, Doktor S, Desino K. Global Analysis of Models for Predicting Human Absorption: QSAR, In Vitro, and Preclinical Models. *J Med Chem*. 2021;64(13):9389–403.
 36. Sathish JG, Bhatt S, DaSilva JK, Flynn D, Jenkinson S, Kalgutkar AS, Liu M, Manickam B, Pinkstaff J, Reagan WJ, Shirai N, Shoieb AM, Sirivelu M, Vispute S, Vitsky A, Walters K, Wisialowski TA, Updyke LW. Comprehensive Nonclinical Safety Assessment of Nirmatrelvir Supporting Timely Development of the SARS-COV-2 Antiviral Therapeutic. Paxlovid Int J Toxicol. 2022;41(4):276–90.
 37. Chiou WL, Chung SM, Wu TC, Ma C. A comprehensive account on the role of efflux transporters in the gastrointestinal absorption of 13 commonly used substrate drugs in humans. *Int J Clin Pharmacol Ther*. 2001;39(3):93–101.
 38. Lee YJ, Chung SJ, Shim CK. Limited role of P-glycoprotein in the intestinal absorption of cyclosporin A. *Biol Pharm Bull*. 2005;28(4):760–3.

39. Cao X, Yu LX, Barbaciru C, Landowski CP, Shin HC, Gibbs S, Miller HA, Amidon GL, Sun D. Permeability dominates in vivo intestinal absorption of P-gp substrate with high solubility and high permeability. *Mol Pharm*. 2005;2(4):329–40.
40. Berggren S, Hoogstraate J, Fagerholm U, Lennernäs H. Characterization of jejunal absorption and apical efflux of ropivacaine, lidocaine and bupivacaine in the rat using in situ and in vitro absorption models. *Eur J Pharm Sci*. 2004;21(4):553–60.
41. Chiou WL, Chung SM, Wu TC. Apparent lack of effect of P-glycoprotein on the gastrointestinal absorption of a substrate, tacrolimus, in normal mice. *Pharm Res*. 2000;17(2):205–8.
42. Lin JH. How significant is the role of P-glycoprotein in drug absorption and brain uptake? *Drugs Today (Barcelona, Spain: 1998)*. 2004;40(1):5–22.
43. Toyobuku H, Tamai I, Ueno K, Tsuji A. Limited influence of P-glycoprotein on small-intestinal absorption of cilostazol, a high absorptive permeability drug. *J Pharm Sci*. 2003;92(11):2249–59.
44. Bentz J, O'Connor MP, Bednarczyk D, Coleman J, Lee C, Palm J, Pak YA, Perloff ES, Reyner E, Balimane P, Brännström M, Chu X, Funk C, Guo A, Hanna I, Herédi-Szabó K, Hillgren K, Li L, Hollnack-Pusch E, Jamei M, Lin X, Mason AK, Neuhoff S, Patel A, Podila L, Plise E, Rajaraman G, Salphati L, Sands E, Taub ME, Taur JS, Weitz D, Wortelboer HM, Xia CQ, Xiao G, Yabut J, Yamagata T, Zhang L, Ellens H. Variability in P-glycoprotein inhibitory potency (IC₅₀) using various in vitro experimental systems: implications for universal digoxin drug-drug interaction risk assessment decision criteria. *Drug Metab Dispos*. 2013;41(7):1347–66.
45. Cook JA, Feng B, Fenner KS, Kempshall S, Liu R, Rotter C, Smith DA, Troutman MD, Ullah M, Lee CA. Refining the In Vitro and In Vivo Critical Parameters for P-Glycoprotein, [I]/IC₅₀ and [I₂]/IC₅₀, That Allow for the Exclusion of Drug Candidates from Clinical Digoxin Interaction Studies. *Mol Pharm*. 2010;7(2):398–411.
46. Izumi S, Nozaki Y, Maeda K, Komori T, Takenaka O, Kusuhara H, Sugiyama Y. Investigation of the impact of substrate selection on in vitro organic anion transporting polypeptide 1B1 inhibition profiles for the prediction of drug-drug interactions. *Drug Metab Dispos*. 2015;43(2):235–47.
47. Yabut J, Houle R, Wang S, Liaw A, Katwaru R, Collier H, Hittle L, Chu X. Selection of an Optimal In Vitro Model to Assess P-gp Inhibition: Comparison of Vesicular and Bidirectional Transcellular Transport Inhibition Assays. *Drug Metab Dispos*. 2022;50(7):909–22.
48. Yoshida K, Guo C, Sane R. Quantitative Prediction of OATP-Mediated Drug-Drug Interactions With Model-Based Analysis of Endogenous Biomarker Kinetics. *CPT: Pharmacometrics Syst Pharmacol*. 2018;7(8):517–24.
49. Li R, Barton HA, Varma MV. Prediction of Pharmacokinetics and Drug-Drug Interactions When Hepatic Transporters are Involved. *Clin Pharmacokinet*. 2014;53(8):659–78.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.